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The Biornimetic Synthesis of Marine Alkaloid Related Fyrido- and Pymolo[2,3,4-kl]acridines

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Abstract: A biomimetic reaction between β , β' -diaminoketones (e.g. kynuramine, kynurenine or 0,0'-diaminobenzophenone) and a variety of cyclohexanediones and quinones leading to pyrido[2,3,4-kl)acridines is described. The synthesis of several di- and tetrahydro--pyrido[2,3,4-kl)acridine derivatives (7, 10, 12 and 15) as well as benzoderivatives of the marine alkaloids eilatin (1) and ascididemin (2), compounds 28 and 30, has been **accomplished. Additionally, the new hetemcycles isoeilatin (24), and diazapmtacem 19 have also been syndmized. All newly prepamd hetemcycles have ban fully characterized** by IR, MS and mainly by NMR spectroscopy. An analogous synthesis has been developed fo pyrrolo[2,3,4-*kI*) acridines, the heterocyclic core of the bioactive plakinidines (31 A-C).

The structures of over 40 pyrido[2,3,4-k*I*]acridine alkaloids have been published over the last decade¹. This group of alkaloids is currently of great interest due to their significant biological activities¹. Almost all pyridoacridines have been reported as having significant cytotoxity¹. Other activities such as the regulation of cellular growth and differentiation, affect on cAMP-mediated processes², inhibition of topoisomerase II, and anti-HIV activity¹ have also been reported. Our discovery of six compounds of this group of alkaloids (eilatin, debromoshermilamine and the segolins) which possess interesting biological activities⁴³ brought us to the synthesis of pyrido^{[2,3,4-kl]acridines.}

Recently we reported a novel biomimetic synthesis of pyrido[23,4-Mlacridines by the reaction of kynuramine derivatives with a variety of quinones, hydroquinones and dienes^{4,5}. Using this synthesis

S-ME 1: **Suggested biogenesis for 1 and 2**

we synthesized the marine alkaloids eilatin⁵ (1) and ascididemin⁶ (2). The existence of kynuramine and quinolinequinones in marine organisms^{7,8,9} suggests that the title biomimetic reaction may also describe the biosynthesis of eilatin (1), ascididemin (2) (Scheme 1). and other pyrido[2,3,4-kl]acridine alkaloids such as biemnadin¹⁰ (3), as demonstrated in Scheme 2. The latter proposed biogenesis is supported by the fact that in the Biemna sponge biemnadin is accompanied by its suggested precursor 8,9-dihydro-11-hydroxyascididemin (4) which itself may be obtained from quinolinequinone 5 and kynuramine (6) (Scheme 2). Compound 4 might react sequentially with a second molecule of kynuramine (6) to give biemnadin (3) via intermediates m and n by an aldol addition reaction followed by a Michael reaction (Scheme 2). Similar biogeneses can also be suggested for eudistones A and $B¹¹$ and other less complicated members of this pyridoacridine group¹.

SCHEME 2: Suggested biogenesis for biemnadin

We have demonstrated that 1,3-cyclohexanedione reacts with kynuramine (6) under acidic oxidative conditions to give dihydropyrido[2,3,4-kI]acridine 7 which oxidizes further to pyridoacridine 8^4 . We show here that kynurenine (9) (the biogenetic precursor of kynuramine which retains the carboxylic group), its acetamido methyl ester derivative 11 (Scheme 5), and the N-trifluoroacetamido-3'-methyleneacetoxy derivative of kynuramine $(13)^{12}$ (Scheme 5) also give the di- or tetrahydropyridoacridine systems, namely compounds 10, 12, and compound 15 *via* 14, respectively. Each of these three compounds (10, 12, 15) was fully characterized spectroscopically mainly by 1D and 2D-NMR experiments (COSY, TOCSY, d-NOE, HMQC and HMBC correlations). Both compounds 10 and 12 undergo decarboxylation under stronger acidic conditions (acetic acid - HCl; $20:1$) to give compound 7 (Scheme 3). The reaction of 1,3-cyclohexanedione with the kynuramine derivative 13 afforded the dihydroacridanone 14 by the Friedländer quinoline synthesis¹³ (via a Schiff base followed by condensation between the carbonyl of the kynuramine moiety and the active methylene of the cyclohexanedione) (Scheme 3). The IR and NMR data of 14 (Experimental) clearly point to the formation of the quinoline and to a cyclohexanone

group, fused to this quinoline. Treatment of 14 with ammonia afforded compound 15- the 2-methylene-**-oxy derivative of compound 7. Compounds 10, 12 and 15 may serve as starting materials for the preparation of C2 and/or N3 substituted pyridoacridines.**

The reaction of 1,3-cyclohexanedione with N-trifluoroacetamidokynuramine (17a)⁶ in acetic acid for 1h afforded, as with 14, a dihydroacridanone derivative (compound 18a, Scheme 4). The IR and NMR data of 18a (Experimental) secured the remaining of one carbonyl group at C1, conjugated with the **quinoline. As with 14, treatment of dihydroacridanone 18a with ammonia afforded compound 7. By the** same route the kynurenine derivative 17b afforded compound 10 via compound 18b (Scheme 5).

Nitration of compound 7 with nitric acid at 100°C gave one major mono nitro derivative (16) (Scheme 3). From the proton NMR spectrum [one low field aromatic proton $(\delta 9.43d, J=1.8Hz; H-11)$ and **an AB spin system (8 8.6Odd, J=8,1.8Hz, H-9 and 8 8.23d, J=8Hz. H-8)], it was clear that the nitration** took place on the benzene ring either at C9 or C10. As the doublet of the proton α to the nitro **substituent (8 9.43) exhibited an NOB with H-l (4%). it was evident that the nitration took place on** C10. Compound 16 is expected to be another good starting material for all kinds of C10-substituted pyrido^{[2,3,4-k]acridines via its diazonium derivative.}

Reacting 1.4-cyclohexanedione with compounds 17a or 17b, gave the 1:2 adducts, namely compounds 19a or 19b, respectively. The ¹³C NMR spectrum of both 19a and 19b, which demonstrated only ten carbon resonances for the pentacyclic system and only one set of resonances for the two ethyltrifluoroacetamide side chains, pointed clearly to a symmetric molecule. Careful interpretation of the NMR spectra of compound 19a (Experimental), determined the 1,7-disubstituted-6,12-diazapentacene structure (Scheme 4). The suggested linear heterocycle was preferred over the angular dibenzo-4,7phenanthroline isomer on the basis of an NOE between $H-1'(1[*])$ and $H-13(14)$, a spatial proximity which is impossible in the case of the angular structure where the two ethylfluoroacetamido functionalities at positions 1 and 10 of the 4,7-phenanthroline are on the opposite side of the C5,C6 ethylene.

SCHEME 5

Interestingly, in contrast to the carbon NMR spectrum, the proton spectrum of 19a differentiated between the geminal protons of the various methylenes as is evident from the COSY, HMOC and HMBC experiments. The first experiment determined the geminal pairs, the second confirmed the pairs and determined the carbon resonances of the atoms carrying each pair, and the HMBC experiment gave the correlations between the protons and their neighboring C-atoms. The proton resonances suggest an asymmetric conformation in 19a, maybe due to strong hydrogen bonds which are responsible for the geminal methylene protons not being equivalent.

Following the study of the 1.3- and 1.4-cyclohexanedione isomers the investigation of the reaction between 1,2-cyclohexanedione and the kynuramine derivative $17a^6$ was undertaken. Mild conditions (reflux for 1h in CH₃CO₂H and catalytic amounts of HCl) afforded a 1:1 adduct, 20. The spectral data of 20 (Experimental) suggested a single Friedländer quinoline synthesis product (Scheme 4). Compound 20 might be a promising precursor for other substituted benzophenanthrolines¹⁴.

Once compounds 19a and 19b were obtained, the preparation of a 13,14-dioxo derivative as a potential intermediate to isocilatin (24) (dibenzo[l,k]-1,6,7,12-tetraazaperylene) was investigated. For this purpose the reaction between 2,5-dihydroxy-1,4-cyclohexanedione (21) and compound 17a, under acidic conditions, was performed. Indeed, the latter reaction afforded a 2:1, kynuramine-quinone adduct (23) (Scheme 6). In contrast to compounds 19a and 19b, this adduct was not symmetric according to its spectral data. On the basis of its NMR data (Experimental) compound 23, has been shown to be the 2-kynuramino-p-acridine quinone. The H-3 singlet at δ 6.65 in the NMR spectrum was characteristic of the suggested structure as were the resonances of the kynuramine moiety (Experimental). Tindale¹⁵ reported the reaction of compound 21 with several amines in ethanol to give the 1:2 "tetrol" adducts of the general structure 22 (Scheme 6). When compound 21 was reacted with 17a in ethanol rather than in a CH₃CO₂H/ TFA mixture a similar tetrol (22) was obtained, as characterized by its proton NMR spectrum [δ 7.51 (d, J=8Hz,1H), 7.32 (d, 8Hz,1H), δ 7.18 (t, J=8Hz,1H), 7.13 (t, J=8Hz,1H), 7.02 (s, 1H: the center tetrol ring proton), 3.68 (q, J=6Hz, 2H) and 3.00 (t, J=6Hz, 2H)]. The symmetric structure of 22 supports the position of the C-N bonds of 23 and hence the structure of 24.

12964

As demonstrated earlier^{4,5} compound 23 could be expected to form two pyridine rings by treatment with ammonia. Indeed, the ammoniacal treatment of 23 afforded a heptacyclic compound, called isoeilatin (24) (Scheme 6). The structure of 24 was fully characterized by NMR data (Experimental), including COSY, d-NOE (between H-1(9) and H-16(8)), HMQC and HMBC correlations. Whereas the proton NMR data of 24 were similar to those of eilatin³ (1), the ¹³C data in the same solvent (CDCl_/TFA; 10:1) were completely different:

1:8 147.3d, 144.1s, 142.9s, 142.3s, 141.6s, 136.7s, 134.1d, 128.9d, 125.0d, 123.0d, 121.5d, 116.9s, 24: δ 146.4s, 143.7s, 143.1s, 141.3d, 136.8d, 133.9d, 132.9d, 131.5s, 125.2d, 121.9d, 121.3s, 117.0s.

The synthesis of two additional derivatives of marine pyrido[2,3,4-kl]acridine alkaloids (compounds 28 and 30) is shown in Scheme 7. Benzoascididemin (28) and dibenzoeilatin (30) were prepared employing the title synthesis using 2.2'-diaminobenzophenone⁴ (26) and quinolinequinone⁶ 25 or o-benzoquinone¹⁶, respectively (Scheme 7). The first synthesis resembles the synthesis of ascididemin⁶, using 2,2'-diaminobenzophenone (26) instead of kynumarine. Comparison of the NMR data of 27 and benzoascididemin (28) with those of ascididemin and its synthetic precursor⁶, fully supported the The bioactivity of the latter two compounds is presently under structure of both 27 and 28. investigation.

In an earlier report⁴ we described the synthesis of a quinolinoacridanone ("benzopyrido-[2,3,4-kl]--acridine"), the "right" half of 29 (Scheme 7), from o-benzoquinone and 2.2'-diaminobenzophenone (26). In that report⁴ we also mentioned the formation of an additional 1:2 adduct which has now been obtained in somewhat better yield (36%). The spectroscopic data (Experimental) of this 1:2 adduct, compound 29, had much in common with that of the above mentioned, earlier reported, 1:1 adduct, quinolinoacridanone⁴, and with that of 2,2'-diaminobenzophenone (26), these being the two halves of 29.

Acidic treatment of compound 29 (CH₃CO₂H/H₂SO₄/ TFA; 45:10:45) at 60^oC for 40 min, afforded a highly symmetrical compound (30), $C_{32}H_{16}N_A$, as judged from the HRMS and carbon NMR spectrum. Only

nine aromatic resonances were observable in the carbon spectrum and the proton NMR spectrum showed even fewer signals, i.e. only four protons $(8\quad9.32, 8.85, 8.35, 8.23$ ppm), both suggesting the symmetrical nonacyclic dibenzoeilatin structure 30 (tetrabenzo[b,e,k,n]-1,6,7,12-tetraazaperylene) (Scheme 7). The latter structure gives a good explanation for the very low resonauce of four of the protons (6 9.32) because of the very crowded structure (overlapping of H-l (and H-10) and H-20 (and H-11)). It is suggested that the entire molecule of 30 is twisted to a certain degree around the ClOb, C15c, C5b, C2Ob axis due to the expected repulsions between the aforementioned protons.

Recently, in addition to the above discussed pyrido $[2.3,4-k/l$ acridine alkaloids, three novel bioactive pyrrolo[2,3,4- k]acridines, plakinidines A-C (31) have been isolated from the marine sponge Plakortis sp.^{17a,b} These pentacyclic aromatic alkaloids belong to a new class which has not yet been reported from marine or terrestrial origin. Here we suggest a biogenesis for these alkaloids similar to the one proposed for the pyridoacridines, namely a reaction between the lower homologue of kynuramine, 2, β -diaminoacetophenone (32a), and an appropriate quinoline (33) (Scheme 8). As mentioned **ahove** oxygenated quinolines like trihydroxyquinoline 34 have previously been isolated from marine sources⁹. To support this postulate we undertook the synthesis of a pyrrolo^{[2}.3,4- k] acridine through a condensation of 13-cyclohexauedione with 2-amino-2'-acetamido-acetophenone **32b. This reaction** afforded ca. 50% yield of the desired tetrahydro pyrroloacridine 35, $C_{14}H_{12}N_2$. The structure of 35 was unequivocally confirmed by its NMR spectra (Experimental), namely correlations between H-1 and C2b, 6a, 5a, lob; H-7 with ClOa; H-5 with C2b, 5a and H-10 with ClOb, 9,6a as observed in an HMBC experiment, and an 8% NOE between H-1 and H-10. Compound 35 is relatively unstable and so far we have not found the right oxidation conditions to transform it to the pyrroloacridine itself.

The motivation for searching for new marine natural products as potential drugs is that once biologicslly interestmg metabolites are discovered, they, or related compounds, will be synthesized in the laboratory. The above described biomimetic syntheses are very good examples for this concept and indeed several of the newly synthesized compounds which are presently under investigation show interesting bioactivities.

EXPERIMENTAL

IR spectra were recorded on a **Nicolet 205 R-IR spectrophotometer. Low resolution mass spectra** were recorded on a Finnigan-4021 mass spectrometer. ¹H and ¹³C NMR spectra were recorded on Bruker AMX-360 and ARX-500 spectrometers. All chemical shifts are reported with respect to TMS (8=0). HRMS **were taken on a** JEOL SX 102 instroment. One and multi bond CH-correlations were established by HMQC and HMBC experiments, respectively, and given as C/H's. Protons locations were determined by COSY (through bond) and by d-NOE (through space) correlations. All chromatographics were carried out by vaccum liquid chromatography on silica-gel H and TLC on silica-gel (Merck 7735).

Dihydropyrido[2,3,4-kl]acridine (7).

Method A: Compound 10 (or 12) (32mg, 0.1mmol) was refluxed in a 20:1 mixture of CH₃CO₂H-HCl (10 mL) for 1h. The mixture was poured over ice (20g), brought to pH 8 with conc. NH₃ and then extracted with CH_aCl_a (3x20mL). After evaporation of the solvent the residue was purified chromatographically (eluted with CH₂Cl₂) to give 7 as a yellow oil (13mg, 58%) identical to the earlier reported compound.⁴

Method B: Compound 17a (34mg, 0.1mmol) was stirred for 24h in a solution of 28% ammonia (0.5mL) in MeOH (5mL). Purification of the residue, after evaporation of the ammoniacal methanol solution, by chromatography afforded compound 7^4 (14mg, 64%).

2-Carboxy-dihydropyrido[2,3,4-kl]acridine (10).

Method A: Kynurenine (21mg, O.lmmol), 1,3-cyclohexanedione (13mg, O.llmmol) and sodium m-nitrophenylsulfonate (45mg, 0.2mmol) in CH₂CO₂H (5mL) were refluxed for 1h. Work-up, as described for 7 (method A), afforded compound 10 (13mg, 50%); yellow powder, mp. 168° . MS: $m/z(\%) = 264 \text{ (M}^+,$ $C_{16}H_{12}N_2O_2, 6$, 263(M⁺- H, 33), 262(M⁺-2H, 100), 220(M⁺- CO₂, 4); IR (KBr): v = 3436 bs, 1687, 1604, 1583, 1423, 786 cm⁻¹; ¹H-NMR (5% CD₃OD/CDCl₃): δ 9.17 (s, H-1), 8.61 (d, J=8Hz, H-11), 8.18 (d, J=8Hx, H-8), 6 7.84 (t, J=IHx, H-9), 7.69& J=IHx, H-10). 3,5O(t, J=5.5Hx, 2H-4), 3.37(t, J=SHx. 2H-6), 2.36 (m, 2H-5). ¹³C-NMR (5% CD₃OD/CDCl₃): δ 167.0(s, CO₃H), 161.4(s, C-7a), 160.6(s, C-6a), 146.9(s, C-3a), 144.9 (s, C-2), 138.7(s, C-11a), 131.8(d, C-11), 130.5(d, C-8), 127.2(d, C-9), 123.2(d, C-lo), 122.2(s, C-llb), 119.2(s, C-3b), 113.3(d, C-l), 33.9(t, C-4), 33.6(t, C-6), 22.7(t, C-5). Observed CH-correlations (C to H's): 3a/5,5'; 3b/1,4; 4/5,5',6,6'; 6a/5.5',6,6'; 7a/8,9; 8/9; 9/10,11; 10/8; 11/10; 11a/1, CO₂/1.

Method B: Compound 18b (20mg, 0.05mmol) gave compound 10 (12mg, 60%) under the same ammoniacal**methanol conditions as described for the synthesis of 7 (method** B).

N(3')-Acetamidokynurenine methyl ester (11). A solution of N(2')-acetamidotryptophan methyl ester¹⁸ (2.6g, 10mmol) in CH₃CO₂H (30mL) was treated with ozone for 5 min at r.t.. Conc. HCl (2mL) was then added and the solution was left at r.t. for 24h. The mixture was then poured over ice (5Og), made basic with conc. NH₃ (to pH 8) and then extracted with CH₂Cl₃ (3x50 mL). The combined organic phase was washed with brine, dried over anhy. Na₂SO₄ and evaporated to give a brownish oil (2.21g, 84%), Rf=0.5 (ethyl acetate), which was not further purified; MS: $m/z(\%) = 264 \ (M^{\dagger}, C_{13}H_{16}N_{10}Q_{10}$, 100); IR (KBr): $v = 3341, 1738, 1732, 1658, 1600, 1538, 1216, 987, 850$ cm⁻¹; ¹H-NMR (CDC1):8 7.59(d,J=8, H-3), 7.29(t, J=8, H-5), 6.62(d, J=8, H-6), 6.59(t, J=8, H-4), 6.17(bs, NH), 4.88(m, H-2'), 3.77(dd, J=16,5) and $3.48(\text{dd}, J=16, 5)$) (2H-1[']), $3.62(s, OMe)$, $1.98(s, CH_{s}CO)$.

1,2,5,6-Tetrahydro-3-acetamido-2-carbomethoxypyrido[2,3,4-k]acridine (12). Compound 11 (264mg, 1 mmol), was refluxed together with 1,3-cyclohexanedione (123mg, 1.1 mmol) in CH₃CO₂H (20mL) for 1h.

Work-up, as described for 7 (method A), gave a brown oil which was purified by chtomatography (eluted with 2% MeOH in CH_aCl₂) to afford a brownish oil (145mg, 45%). HRMS (FAB⁺): 323.1397 (Δ mmu+0.2) (MH⁺, C₁₉H₁₉N₂O₃, 100%); MS: m/z (%)=322 (M⁺, C₁₉H₁₈N₂O₃,13%), 321(M⁺- H, 60), 261(M⁺-H-CO₂Me, 10); IR(KBr): v = 1740, 1656, 1433, 1382, 1216, 1097, 995 cm⁻¹; ¹H-NMR (CDCL₂): δ 8.00(d, J=SHx, H-11). 7.93 (d. J=SHx, H-8). 7.7O(t, J=SHz, H-9). 7.57 (t, J=SHx, H-lo), 6.13(bs, H-2), 6.06&s, H-4). 3.% (d, J=17Hx, H-l), 3.68(s, OMe), 3.37(dd, J=17,2Hx, H-l'), 3.10 (m, 286), 2.62(m. 2H-5), 2.39(s, COCH₃); ¹³C-NMR (CDCl₃): δ 170.5(s, CO₂Me), 169.4(s,COCH₃), 156.7(s, C-6a), 146.3(s, C-7a). 133.7(s. C-lla), 132.1(s, C-3a). 129.3(d, C-10). 129.qd. C-11), 126.4(d, C-9), 125.9(s, C-lib), 123.6(d, C-4). 122.9(d. C-S), 120.9(s, C-3b), 52.S(q. OMe), SO.l(d, C-2), 31.O(t, C-6), 25.9(t, C-1), 22.2(t, C-5), 21.1(q, COCH₂). Observed CH-correlation (C to H's): CO₂Me/1,1'; 3a/5,5'; $3b/1,1', 4/5.5', 6,6'; 6a/5,5', 6,6'; 7a/8,9; 8/9; 9/10, 11; 10/8; 11/10; 11a/1,1',8; \text{CO}_2CH_4/CO_2CH_4.$

o-Aminophenyl 2-trifluoroacetamido-3-acetylpropyl ketone (13).

a. N(2')-Trifluoroacetamidotryptophanol. L-Triptophanol¹² (1.88g, 10mmol) was stirred in methanol (40mL) with $CF₁CO₂Et$ (2.84g, 20mmol) and $NEt₁(2g, 20mmol)$ for 1h at r.t.. The methanol was then removed under vacuum, the residue taken up into CH₂Cl₂ (50mL) washed with 5% aq. NH₃ and then dried over anhy. Na₂SO₄ to give, after evaporation of the CH₂Cl₂ a yellow oil (2.6g, 90%), Rf=0.5 (CH₂Cl₂); MS: $m/z(\%) = 287 (M^+ + H, C_{13}H_{14}N_2O_3F_3, 73)$, 286 (M^F,70); IR(KBr): v = 3386, 1674, 1613, 1522, 1343, 1107, 760 cm⁻¹; ¹H-NMR (CDCl₃): δ 7.62 (d, J=8, H-4), 7.31(d, J=8, H-7), 7.13(t, J=8, H-6), 7.08(t, J=8, H-S), 7.02(s, H-2), 4.23(m, H-2'). 3.57(dd, J=S.l; 2H-3'). 3.01(d, J=7. 2H-1'). The oil was used in the next step without further purification.

b. N(2')-Trifluoroacetamidotryptophanol acetate. The above oil (2.87g. lOmmo1) was stirred overnight at r.t. in a 1:l mixture of pyridine-acetic anhydride (10 mL). Vacuum distillation of the excess pyridine-acetic anhydride mixture left a yellow oil (3.288, 100%) which was used in the next step without further purification, Rf=0.6 (EtOAc); MS: $m/z(\%)$ = 329 (M⁺+ H, C₁₄H₁₆N₂O₃F₃, 23), 328 (M⁺, 81); IR (KBr): $v = 3282$, 1713, 1682, 1583, 1466, 1238, 1030, 820 cm⁻¹; ¹H-NMR (CDCI_a): δ 7.51(d, J=8, H-4). 7.26(d. J=8, H-7). 7.08(m. 2H-5.6). 6.93(s, H-2), 4.46(m, H-2'), 4.09(dd, *J=S,* 2; 2H-3'), 2.94(d, $J=7$, 2H-1'), 1.98(s, CH₂CO).

C. N(3')-Trifhtoroacetamidokynurinol acetate. Oxonolysis of the above oil (3.28g. 10 mmol) under the same conditions as described for the synthesis of 11 gave a yellow oil (1.52g, 45%). Rf=0.8 (CH₂Cl₂-EtOAc; 1:1); MS: $m/z(\%) = 332 (M^+, C_{14}H_{15}N_2F_3O_4, 71)$; IR (KBr): v= 3178, 1719, 1706, 1674, 1600, 1339, 1061, 910 cm⁻¹; ¹H-NMR (CDCl₄): δ 7.54 (d, J=8, H-3), 7.21(t, J=8, H-5), 6.60(d, J=8, H-6), 6.56(t, $J=8$, H-4), 4.61(m, H-2'), 4.28 (dd, $J=10,5$) and 4.20 (dd, $J=10,5$) (2H-3'), 3.29 (dd, $J=16,5$) and 3.18 (dd, $J=16,5$) (2H-1[']), 1.97 (s, CH₂CO).

3,4-Dihgdro-9(2'-trilluowwetam iaO-3'-a~~ropyl)-l-a~i~one (14). Compound 13 (33mg, 0.1 mrnol). and 1,3-cyclohexanedione (13mg. 0.11 mmol) were refluxed in acetic acid (SmL) in the presence of 1 drop of conc. HCl for 1h. After the same work-up as described for 7 (method A) the residue was chromatographed (eluted with 2% MeOH in CH_2Cl_2) to afford compound 14 as a brownish-yellow oil (16mg,55%), Rf=0.7 (EtOAc); HRMS (FAB)⁺: 409.1384 (Δ mmu + 3.1) (MH⁺, C₂₀H₂₀N₂O₄F₃, 100%); IR (KBr): $v = 3370$, 1723, 1706, 1692, 1601, 1566, 1222, 1127, 935 cm⁻¹; ¹H-NMR (CDCL₃): δ 8.18 (d, J=8Hz, H-S), 8.07 (d, J=SHx, H-Sj, 7.81 (t, J=SHz, H-6). 7.63 (t, J=SHx, H-7). 4.62 (m, H-2'), 4.41 (m, 2H-CH_2OAc), 3.8S(t, J=lOHx) and 3.SSm (2H-l), 3.31 (t, J=6Hx, 2H-4), 2.90 (t, *J=6Hz,* 2H-2). 2.22 (m, 2H-3), 2.18 (s, CH₄CO); ¹³C-NMR (CDCl₄): δ 203.0(s,C-1), 170.0 (s, OAc), 161.5 (s,C-9), 148.0 (s. C-8b). 146.0(s, C-4a), 132.5 (d, C-6). 129.0 (d, C-5). 127.1 (d, C-7). 126.1(s, C-4b), 124.2 (d, C-8), 113.9 (s, C-8a), 65.2 (t, CH₂OAc), 51.2 (d, C-2'), 40.3 (t, C-1'), 34.1 (t, C-4), 29.2 (t, C-5). 21.6 (t, C-6), 20.0 (q, CH₃CO). Observed CH-correlations (C to H's): $1/2$; OCOCH₁/3'; 4a/1',1"; 4b/4,1',1"; 5/7; 6/8; 7/5; 8b/6,8.

4,5-Dihydro-2-methylenehydroxypyrido[2,3,4-kl]acridine (15). Compound 14 (21mg, 0.05mmol) was treated with $NH₃$ under the same conditions as described for the synthesis of 7 (method B) to provide after chromatography (eluted with EtOAc/CH₂Cl₃; 1:1) compound 15 (13mg, 55%.) as a yellow oil, Rf = 0.4 (EtOAc); HRMS (FAB⁺): 251.1176 (Δ mmu - 0.8) (MH⁺, C₁₆H₁₅N₂O, 100%); IR (KBr): v = 3345, 2961, d_z-1613, 1477, 1209, 863 cm⁻¹; ¹H-NMR (d₆-DMSO): δ 8.69 (d_p \overrightarrow{d} = 8Hz, H-11), 8.51 (s, H-1), 8.03 (d_p J = 8Hz, H-8), 7.84 (t, J = 8Hz, H-9), 7.69 (t, J = 8Hz, H-10), 4.75 (s, CH₂OH), 3.22 (m,4H-4,6), 2.18 (m, 2H-5); ¹³C-NMR (CDCl₃): δ 161.7 (s,C-7a), 160.9 (s,C-6a), 158.5 (s,C-2), 144.9 (s,C-3a), 138.9 $(s, C-11a), 131.0 (d, C-11), 128.9 (d, C-8), 126.6 (d, C-9), 122.9 (d, C-10), 121.9 (s, C-11b), 117.3$ $(s, C-3b)$, 109.9 (d,C-1), 64.9 (t, CH₂OH), 34.2 (t,C-4), 33.5 (t,C-6), 22.8 (t,C-5).

Dihydro-10-nitropyrido $[2,3,4-k]$ acridine (16). Compound 7 (22mg, 0.1mmol) in a mixture of fuming HNO₃-conc. H₂SO₄ (1:1) (5mL) was heated for 1h at 100^oC. The mixture was cooled, poured over ice (10g), and conc. NH₃ (20mL) was added. The mixture was then extracted with CH₂Cl₃(3x 20mL), the organic phase was washed with brine, dried over anhy. Na₂SO₄ and evaporated to give a dark red oil (20mg). Chromatography of the crude oil (eluted with 5% MeOH in CH₂Cl₂) gave pure 16 (14mg, 54%), as a yellowish powder, m.p. 197°, Rf=0.5 (EtOAc); HRMS (FAB⁺): 266.0925 (Δ mmu-0.5) (MH⁺, C₁₅H₁₂N₃O₂, 100); MS: $m/z(\%) = 265 (M^+, C_{15}H_{11}N_3O_2,100)$, 235(M⁺- NO,11); IR (KBr): v = 1634, 1588, 1386, 1172, 1008, 910 cm⁻¹; ¹H-NMR (5% CD₃OD/CDCl₃): δ 9.43(d, J=1.8Hz, H-11), 8.88 (d, J=5.6Hz, H-2), 8.60 (dd, J=8, 1.8Hz, H-9), 8.36 (d, J=5.6Hx, H-l), 8.23(d, J=8Hx, H-S), 3.38 (m, 4H-4.6). 2.39 (m, 285). An NOE of 4% was measured between H-1 and H-11.

3,4-Dihydro-9-(2'-trifluoroacetamidoethyl)-1-acridanone (18a). Compound 17a⁶ (26mg, 0.1mmol) and 1,3cyclohexanedione (13mg. O.llmmol) were refluxed for lh in acetic acid (5mL). After work-up as described for compound 7, the product was purified by chromatography (eluted with $CH_{A}Cl_{A}$) to afford compound 18a (22mg, 61%) as a yellow oil, Rf=0.8 (EtOAc); MS: m/z (%)=337 (M⁺+ H, C₁₂H₁₆P₃N₂O₂,19), 336 (M⁺, 100), 224 (M⁺- NHCOCF₃, 20), 223 (M⁺- NHCOCF₃-H, 98); IR (KBr): v = 3112, 2941, 1718, 1681, 1561, 1213, 1183, 1156, 784 cm⁻¹; ^{'1}H-NMR (CDCl_a): δ 8.25(d, J=8Hz, H-8), 7.99(d, J=8Hz, H-5), 7.75(t, J=8Hx, H-6), 7.570, J=8 Hz. H-7). 3.74 (m, 4H-,1',2'), 3.27 (t, *J=6 Hz,* 2H-4). 2.83 (t. J=6 Hz, 2H-2), 2.23 (m, 2H-3).

N(3')-trifluoroacetamidokynurenine methyl ester (17b).

a. N-Trifluoroacetamidotryptophan methyl ester. N-Trifluoroacetamidotryptophan methyl ester was synthesized from triptophan methyl ester (2.18g, 10mmol), ethyl trifluoroacetate (2.84g, 20mmol) and NEt₃ (2g, 20mmol) using the same procedure as for compound 13; yellow oil, Rf=0.9 (EtOAc); MS: mlz %) $= 314 \, (\text{M}^+, \text{C}_{14}H_{13}N_0\text{Q}_4F_3,97); \text{ IR (KBr): } v = 3132,1722,1680,1602,1514,1302,1279,1116,863 \text{ cm}^{-1};$ 1 H-NMR (CDCl_a): δ 8.17(bs, NH), 7.39 (d, J=8Hz, H-4), 7.30(d, J=8Hz, H-7), 7.10(t, J=8Hz, H-6), 7.04(t, J=8Hx, H-5), 6.91 (d, J=2Hx, H-2), 4.82 (m, H-23, 3.67 (s, OMe), 3.35 (d. J=SHx, 2H-1'). The compound was used without further purification.

b. N(3')trifluoroacetamidokynurenine methyl ester (17b). Compound 17b was prepared by ozonolysis of

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the above compound using the same procedure as described for the preparation of 13. Compound 17h was obtained as a brown oil, Rf=0.6 (EtOAc). HRMS (FAB⁺): 318.0819 (Ammu-0.8) (MH⁺, C₁₃H₁₃O₄N₂F₃, 100); IR(KBr): 2219, 1724, 1711, 1683, 1600, 1513, 1398, 1255, 1008, 720 cm⁻¹; ¹H-NMR (CDCL): **6** 7.75(d. J=8Hx, H-3). 7.30& J=8Hx, H-5). 6.79(d, J=IHx, H-6), 6.74& 1=8Hz, H-4). 6.37 (bs, NH), 5.03 (m, H-3^o), 3.99 (dd, J=16,5Hz) and 3.61 (dd, J=16,5Hz) (2H-2^o), 3.67 (s, OMe).

3,4-Dihydro-9-(2'-trifluoroacetamido-2'-carbomethoxyethyl)-1-acridanone (18b). Compound 17b (32mg, 0.1mmol) was treated with 1,3-cyclohexanedione (13mg, 0.1mmol) in the same manner as described for the synthesis of 18a, to afford compound 18b (23mg, 58%) as a yellowish oil, Rf=0.6 (EtOAc);

MS: m/z(%) = 394 (M', C₁₉H_{1,}F₃N₁O₄,32), 282 (M'- NHCOCF₃, 19), 281 (M^T- H-NHCOCF₃, 100); IR (KBr): v = 3012, 2986, 1724, 1719, 1658, 1213, 1178, 721 cm⁻t; 'H-NMR (CDCl_{..}): δ 8.29(d, J=8Hz, H-8), 8.04(d, J=8Hz, H-5), 7.80(t, J=8Hz, H-6), 7.63(t, J=8Hz, H-7), 4.91 (m, H-2²), 4.04 (d, J=10 Hz) and 3.88 (d, J=10 Hz) (2H-1'), 3.82 (s, OMe), 3.34 (m, 2H-4), 2.82 (m, 2H-2), 2.23 (m, 2H-3).

1,7-Di(β -trifluoroacetamidoethyl)-6,12-diazapentacene (19a). Compound 17a (52mg, 0.2mmol) and 1,4-cyclohexanedione (12mg, 0.1mmol) were refluxed in CH₃CO₂H (10mL) with catalytic amounts of p-TsOH acid for lh. After work-up as described for compound 7 (method A) the residual browu oil was chromatographed (eluted with ethyl acetate/CH₂Cl₂, 1:1) to afford compound 19a (29mg, 52%), a yellowish oil, Rf=0.7 (EtOAc); HRMS (FAB⁺) 561.1722 (Δ mmu-0.3) (MH⁺, C₂₈H₂₃F₆N₄O₂, 100); MS: $m/z(\%)$: 561 (M⁺+H, C₂₈H₂₃F₆N₄O₂, 14), 560 (M⁺, 41); IR (KBr): v = 1713, 1986, 1402, 1205, 1076, 750 cm⁻¹; ¹H-NMR (CDCL_a): δ 8.20 (d, J=8Hz, 2H-2,8), 8.08 (d, J=8Hz, 2H-5,11), 7.70 (t, J=8Hz, H-4,10), 7.60 (t, J=8Hz, H-3,9), 3.62(m) and 2.80(m) (4H-2 β , 2 β), 3.35(d, J=10Hz, 2H-13,14), 3.00 (d=10 Hz, $2H-13',14'$), $370(m)$ and $3.08(m)$ (4H-2 α ,2 α'); ¹³C-NMR (CDCl_a): δ 161.0 (s, 2C-6a,12a), 157.1 (q, 2COCF₃), 146.0 (s, 2C-5a,11a), 141.0 (s, 2C-1,7), 130.1 (d, 2C-4,10), 128.2 (d, 2C-5, 11), 126.8 (d, $2C-3,9$), 126.2 (s, $2C-13a,14a$), 125.7 (s, $2C-1a,7a$), 124.1 (d, $2C-2,8$), 115.0 (q, $2CF_3$), 40.1 (t, $2C-\beta,\beta'$), 33.2 (t, 2C-13,14), 28.1 (t, $2C-\alpha,\alpha'$). Observed CH-correlations: (C to H's): $1/2$, α,α' ; $1a/3$; $3/5$; $4/2$, 3 ; $5/3$; $5a/2, 4$; $6a/13$, $13'$, $14,14'$; $13a/13,13'$, α, α' .

1,7-Di(β-trifluoroacetamido-β-carbomethoxyethyl)-6,12-diazapentacene (19b). Compound 19b was synthesized from 17b (63mg, 0.2mmol) and 1,4-cyclohexanedione (12mg, 0.01mmol) in the same manner as described for **19a** (31mg, 47%). A brownish oil, Rf=0.7 (EtOAc); HRMS (FAB⁺) 677.1853 (Δ mmu+1.8) $(MH^{\dagger},C_{32}H_{27}N_4O_6F_6$, 100%); MS: m/z %) = 564 (M⁺-NHCOF₃, 100); IR (KBr): v = 1742, 1724, 1546, 1403, 1217, 1191, 742 cm⁻¹; ¹H-NMR (CDCl_a): 8 8.48 (d, J=8Hz, 2H-2,8), 8.11 (d, J=8Hz, 2H-5,11), 7.79 (t, J=8Hx, 2H-3,9), 7.70 (t, J=8Hx, 284,10), 7.01(d, J=7 Hz, 2H-NH-), 4.7Om (2H+,B), ' 3.96(dd, 3=13, 4 Hz) and 3.21(dd, $J=13,4$ Hz) (4H- α,α') 3.36 (d, $J=10$ Hz) and δ 3.13 (d, $J=10$ Hz) (4H-13,14), 3.21 (s, OMe); ¹³C-NMR (CDCI₃): δ 168.0(s, 2CO₂Me), 161.0(s, 2C-6a,12a), 157.5(q,2 COCF₃), 146.8(s, 2C-5a,lla), 137.8 (s, 2C-1,7), 130.2 (d, 2C-4,10), 129.5 (d, 2C-5.11). 127.2 (d, 2C-3.9). 126.2 (s, 2C-1a,7a), 125.9 (s. 2C-13a,14a), 124.3(d, 2C-2,8), 115.0 (q,2CF₃), 53.2 (d, 2C- β , β ²), 52.9 (q, 2 OMe), 33.9 (t, 2C-13,14), 32.1 (t, 2C- α, α'). Observed CH-correlations (C to H's): $1/2$, α, α' ; $1a/3$, α, α' ; 2/4; 3/5; 4/2,3; 5a/2,4; 6a/13,13',14,14'; \angle OCF₄/ β , \angle O₂Me/ α, β , CO₂CH₃.

3,4-Dihydro-9-(2-trifluoroacetamidoethyl)-4-acridanone (20). Compound 20 was synthesized from 17a **(52mg, 02mmol) and 1,2cyclohexanedione (12mg, O.lmmol) in the same manner as described for 14 (22mg.** 30%). A yellowish oil, Rf = 0.5 (EtOAc); HRMS: (FAB⁺): 337.1162 (Δ mmu + 2.1) (MH⁺, C₁₇H₁₆N₂O₂F₃, 100%); IR (KBr): v = 3223, 3081, 2876, 1700, 1690, 1508, 1173, 785 cm⁻¹; ¹H-NMR (CDCL): δ 8.23 (d, J=8Hz, H-8), 8.11 (d, J=8Hz, H-5), 7.72 (t, J=8Hz, H-6), 7.65 (t, J=8Hz, H-7), 3.57 (q, J=6Hz, 2H-2'), 3.45 (t, J=6Hz, 2H-1'), 3.21 (t, J=6.5Hz, 2H-1), 2.85 (t, J=6.5Hz, 2H-3), 2.22 (m, 2H-2); ¹³C-NMR (CDCI₃): δ 197.6 (s, C-4), 158.0 (q, COCF₃), 146.5s, 142.9s, 134.1s, 131.4d, 130.7s, 129.6d, 129.2d. 128.5s, 122.9d, 39.5t, 38.8t, 27.0t, 26.0t, 21.9t.

3(N)-(β-trifluoroacetamidokynuramine)-9-(2'-trifluoroacetamidoethyl)-acridine-1,4-dione (23).

A mixture of β-trifluoroacetamidokynuramine (210mg, 0.8mmol), 2,5-dihydroxy-1,4-benzoquinone (56mg 0.4mmol), in a mixture of CH₃CO₂H/TFA (20:1) (10 mL) was heated under reflux for 2h. After work up, as **described** for 7 (method A) the residue was chromatographed (eluted with EtGAc) to provide compound 23 (32mg, 7%) as a orange powder, m.p. 287-290°C, Rf= 0.5 (EtOAc); MS: m/z (%): 606 (MH⁺, $C_{2.8}H_{2.0}F_{6.7}N_{4.9}Q_{5}$, 2), 492 (M⁺-(H⁺NH₂COCF₃), 23); IR (KBr): v = 3263, 1715, 1620, 1498, 1473, 1089, 733 cm⁻¹; ¹H-NMR (DMSO-d₂): 8 10.98 (bs, NH), 9.76 (bs, NH), 9.43 (bs, NH), 8.59 (d, J=8Hz, H-8), 8.17 (d, J=8Hz, H-5), 8.08 (d, J=8Hz, H-3'). 7.91 (t, J=8Hx), 7.75 (t, J=8 Hz), 7.63 (m, 2H), 7.24 (m, lH), 6.65 (s, H-3), 3.83 (q, J=6Hz, 2H), 3.59 (q, J=6Hz, 2H), 3.51 (t, J=8 Hz, 2H), 3.37 (t, J=8Hz, 2H). Isoeilatin (24). Compound 23 (20mg, 0.033 mmol) was treated with NH_3 under the same condition as described for the synthesis of 7 (method B), to provide after chromatography (eluted with EtOAc/CH_aCl_a (1:1)) compound 24 (5mg, 43%) as a yellow powder, m.p. > 300°C, Rf=0.3 (10% MeOH/CH₂Cl₂); HRMS: (FAB^+) : 357.1136 (Ammu - 0.4) (MH⁺, C₂₄H₁₃N₁, 100%); IR (KBr): v= 3010, 1590, 1200, 1087 cm⁻¹; 1 H-NMR (10% TFA/CDCI_n): 8 9.39 (d. J=6Hz, 2H-7,15), 9.18 (d. J=6Hz, 2H-8,16), 8.89 (d. J=8Hz, 2H-1.9). 8.62 (d, J=8Hz, 2H-4,12), 8.37 (t, J=8 Hz, 2H-3,11), 8.23 (t, J=8Hz, 2H-2,10); ¹³C-NMR (10% TpA/cDc13): 6 146.4 (s. C-4a), 143.7 (s.C-8a), 143.1 (s, C-5b). 141.3 (d. C-7). 136.8 (d, C-3), 133.9 (d, C-2). 132.9 (d. C-4), 131.5 (s, C-5a), 125.2 (d, C-l). 121.9 (d, C-8). 121.3 (s, C-8b). 117.0 (s, C-5c). Observed CH-correlation (C to H's): 1/3; 2/4; 3/1; 4/2; 4a/1,3; 4b/2,4,16; 5b/7; 5c/16; 7/8. **6-N(o,o'-diaminobenxophenone)-quinoline-5,8-quinone (27).** o,o'-Diaminobenxophenone4 (26) (127mg. 0.6 mmol) was reacted with $5,8$ -quinolinequinone under the same conditions as described for the synthesis of 29. Upon chromatography (eluted with 2% MeOH in ethyl acetate) compound 27 was obtained as a red orange powder, m.p. 184[°] (32mg, 17%). Rf = 0.8 (EtOAc); MS: m/z /%) = 369 (M⁺, $C_{2,5}H_{1,5}N_{1}O_{1,48}$, 333 (M⁺- 2H₁O, 35); IR (KBr): v = 3267, 1703, 1643, 1600, 1486, 1402, 1313, 1188, 1071, 860 cm⁻¹; ¹H-NMR (CDCI₄): δ 9.28 (bs, NH), 8.94 (dd, J=5, 1Hz; H-2), 8.34 (dd, J=8, 1Hz; H-4). 7.55 (da, J=8Hx. 5; H-3). 7.52 (t. J=8Hx, H-4'). 7.46 (d. J=8Hx, H-3') 7.43 (t, /=8Hx, H-5'). 7.32 (d. J=8Hz, H-13'), 7.22 (t, J=8Hz, H-11'), 7.16 (d, J=8Hz, H-6'), 6.64 (s, H-7), 6.63 (d, J=8Hz, H-10'), 6.50 (t, J=8Hz, H-12'); ¹³C-NMR (CDCl₄): 198.0 (s, C-7'), 182.0 (s, C-5), 171.0 (s, C-8), 154.1 (d, C-2). 151.8 (8. C-9'). 148.5 (8. C-4b). 144.8 (s, C-6), 136.8 (s. C-l'), 135.0 (d, C-4). 134.8 (d, C-11'). 134.0 (d, C-13'), 131.5 (d. C-3'), 131.0 (a, C-5'). 132.1 (s, C-2'). 127.5 (s. C-4a), 126.0 (d, C-3), 124.2 (d, C-6'), 122.5 (d, C-4'), 117.3 (s, C-8'), 117.0 (d, C-7), 115.2 (d, C-12'), 104.9 (d, C-10'). Observed CH-cormlations (C to H's): 1'/3'.5'; 2/3,4; 2'/4'; 3/Z; 4a/3; 5/4,7; 5'/3'; 6/7; 6'14'; 7'/3',13'; 4bf2.4.7; 8'/10',12'; 9'/11',13'; 11'/13'; 12'/10'.

Benzo[a]ascididemin (28). Compound 27 (20mg, 0.054mmol) dissolved in a mixture of HOAc/H₂SO₄/TFA (45: 10: 45) (5 mL) **was** heated to 60"C for 40 min. After cooling the reaction mixture it was poured over ice (10mg) and the solution was made basic with conc. $NH₃$ (15 mL). The mixture was extracted with $CH_2Cl_2(3x20 \text{ mL})$, the organic phase was washed with brine, dried over anhy. Na₂SO₄ and evaporated. Chromatography of the residue (eluted with 5% MeOH in EtOAc) afforded compound 28, $(13mg, 65\%)$; amorphous yellow powder, m.p. > 300°C, Rf=0.6 (10% MeOH/CH₂Cl₂); HRMS (FAB⁺): 334.0992 $(\Delta$ mmu+1.1) (MH⁺, C₂₂H₁₂N₃O,100%); IR (KBr): v = 1669, 1623, 1596, 1423, 1317, 1196, 1050, 974, 780 cm⁻¹; ¹H-NMR (5% CD₄OD/CDCl₄): δ 9.29 (dd, J=5.5, 1.5Hz, H-9), 9.21 (d, J=8Hz, H-15), 9.13 (d, J=8Hz, H-l), 8.85 (dd, J=S, 1.5Hz; H-7). 8.75 (d, J=SHz, H-4). 8.72 (d, J=SHz, H-12), 8.05 (t, J=SHz, H-13), 8.00 (t, J=SHz, H-3). 7.97 (t, I=SHz, H-14), 7.90 (t, J=SHz. H-2). 7.71 (dd, J=S, 5.5Hz; H-8); ¹³C-NMR (5% CD₃OD/CDCl₃): δ 182.0 (s, C-6), 155.8 (d, C-9), 152.4 (s, C-10a), 147.3 (s, C-4a), 147.2 (s, C-lla), 147.1 (s. C-5b), 144.2 (s. C-Sa). 136.6 (s, C-15b), 136.5 (d, C-7). 133.1 (d, C-4), 132.5 (d, C-12). 131.3 (d, C-13), 130.5 (d. C-14). 130.0 (s, G6a). 129.2 (d, C-2). 127.4 (d, C-l), 127.3 (d, C-15), 125.7 (d, C-S), 124.3 (s, C-15a), 123.2 (s, C-4b). 116.2 (s. C-lob). Observed CH-correlations (C to H's): 4a/1; 6/7; 6a/8; 7/9; 9/7,8; 10a/7; 11a/15; 15a/1,14; 15b/1,15.

8-N(o,o'-diaminobenzophenone)4-onquinlineP,3,4-kl]acridine (29). A mixture of compound 26 (128mg, 0.6mmol), o-benzoquinone¹⁵ (32mg, 0.3mmol) and CeCl_x \cdot 7H_aO (223mg, 0.6mmol) in ethanol (10 mL) was stirred at r.t. overnight. During this time the color changed from yellow to deep red. Water was then added (40 mL) and the mixture was extracted with 5% MeOH in CH₂Cl₂ (3x30 mL). The combined organic phase was washed with brine, dried (anhy. Na₂SO₂) and evaporated to give after chromatography (eluted with EtOAc-CH₂Cl₂, 1:1) compound 29, (53mg, 36%); an amorphous orange powder, m.p. 263°C; Rf=0.8 (EtOAc); HRMS (FAB⁺): 493.1667 (Δ mmu+0.3) (MH⁺, C₃₂H₂₁N₄O₂,100); IR (KBr): v = 3213, 1652, 1566, 1337, 1194, 870 cm⁻¹; ¹H-NMR (CDCl₂): δ 9.04 (d, J=8Hz, 2H-1,13), 8.65 (d, J=8 Hz, H-4 or lo), 8.37 (d, I=SHz, H-10 or 4). 7.78(m, 4H), 7.47(m, 2H). 7.40 (d, /=SHz. lH), 7.28(m, 2H), 6.85(s, H-7), 6.67 (d, J=8Hz, H-12'), 6.56(t, J=8Hz, H-10'), 6.29(bs, NH); ¹³C-NMR (CDCl_a): δ 186.8 (s, C-7'), 182.1 (s, C-6), 151.4s. 149.1s. 147.8s. 146.5s, 145.9s, 144.89 137.1s, 134.W. 134.4d. 133.4d, 132.5d, 131.8d. 131.5d. 131.ld. 130.5d. 129.54 129.ld. 127.2d. 126.9d, 125.7s, 124.3 (d,2C), 123.8s, 122.7s, 122.2s, 118.3s, 117.0d, 115.5d, 115.3s, 103.8d.

Dibenzoeilatin (30). Compound 29 (50mg, 0.11mmol) was treated with a CH₃CO₂H/ H₂SO₁/ TFA mixture (45:10:45) (5 mL) at 60°C for 40 min. Work-up as described for compound 28, except that the alkalified reaction mixture was filtered, instead of extracted. Compound 30 was obtained as a yellow amorphous powder, m.p. > 300°C; Rf=0.4 (10% MeOH/CH₂Cl₂); HRCIMS (Isobutane) m/z, (%):

457.1448 (Δ mmu - 0.8) (MH^+ , C₃₂H₁₇N₄,100); IR (KBr): v = 1636, 1604, 1523, 1369, 1198, 1002, 890 cm⁻¹; ¹H-NMR (5% TFA/CDCl_a): δ 9.32 (d, J=8Hz, H-1), 8.85 (d, J=8Hz, H-4), 8.35 (t, J=8Hz, H-3), 8.23 (t, J=SHz. H-2); "C-NMR (5% TRVCDCl~): 6 140.3 (s, C-4a), 138.9 (s. C-5a). 131.2 (d, C-2), 134.1 (d, C-3), 127.5 (s, C-lob). 126.5 (d. C-l), 125.6 (d, C-4). 121.7 (s. C-4b). 119.3 (s, C-m). Observed CH-correlations (C to H #): 2/4; 3/1; 4/2,3; 4a/1; 4b/1,4.

Dihydropyrrolo[~,4&lacridine (35). 2'-acetamido-2-aminoacetophenone **(32b) prepared from the** corresponding nitro compound¹⁹ by catalytic hydrogenation over 5% Pt/C in MeOH, at 2 atm. $(87.61 \text{ (d,}$ J=SHz, H-3), 7.26 (t, J=SHz, H-4), 6.68 (d, J=SHz, H-6), 6.64 (t, J=SHz, H-5), 4.67 (d, &I.6 Hz, 2H-2'), 2.08 (s, NHCOCH₃); IR (KBr): v = 3186, 1676, 1612, 1566, 1410, 1311 cm⁻¹; MS: m/z %) = 192, $(M^{\dagger}, C_{i_0}H_{i_1}N_1O_{i_2}100)$ (192mg, 1 mmol) was refluxed with 1,3-cyclohexanedione (123mg, 1.1 mmol) in a mixture of CH₃CO₃H/ HCl (20:1) (20 mL) for 1h. After work-up, as described for compound 7, the crude product was chromatographed (eluted with CH_2Cl_2) to give compound 35 (125 mg, 60%) as an unstable yellow oil; Rf=0.6 (EtOAc); MS: $m/z(\%) = 210 (M^+ + 2H, 1)$, 208 (M⁺, C₁₄H₁₂N₂,28); IR (KBr): v= 3099, l617, 1560, 1460, 1366, 1247, 1016, 966 cm⁻¹; ¹H NMR (CDCl₄): δ 7.99 (d, J=8 Hz, H-10), 7.76(d, J=8Hz,

H-7), 7.44 (s, H-1), 7.35 (t, J=8Hz, H-9), 7.34 (t, J=8Hz, H-8), 2.97 (t, J=6Hz, 2H-5.5'), 2.87 (t, J=6Hz, 2H-3,3'), 2.20 (t, J=6Hz, 2H-4,4'); ¹³C NMR (CDCl): δ 159,1 (s, C-2a), 143.8 (s, C-6a), 128.1 (d, C-7), 128.1 (s, C-5a), 124.8 (d, C-9), 123.9 (d, C-8), 122.9 (d, C-10), 122.8 (s, C-10a), 116.2 (s, C-10b), 115.2 (s, C-2b), 106.4 (d, C-1), 29.7 (t, C-3), 23.8 (t, C-4), 21.4 (t, C-5). Observed CH-correlations (C to H's): 1/NH; 2a/3,3',4,4'; 2b/1,3,3',5,5'; 3/NH; 5a/1,4,4'; 5,5'; 6a/1,8,10; 7/8,9; 9/10; 10a/7; 10b/1,10.

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REFERENCES

- Molinski, T.F.; Chem. Rev. 93, 1825 (1993) and references therein. $1.$
- $2.$ Shochet, N.R.; Rudi, A.; Kashman, Y.; Hod, Y.; El-Maghrabi, M.R.; Spector, I.; J. of Cellular Physiol. 157, 481 (1993).
- $3.$ Rudi, A.; Kashman, Y.; J. Org. Chem. 54, 5331 (1989).
- 4. Gellerman, G.; Rudi, A.; Kashman, Y.; Tetrahedron Lett. 34, 1823 (1993).
- Gellerman, G.; Babad, M.; Kashman, Y.; Tetrahedron Lett. 34, 1827 (1993). 5.
- Gellerman, G.; Rudi, A.; Kashman, Y.; Syn. 239 (1994). 6.
- Kashman, Y.; Carmely, S.; Blasberger, D.; Green, D.; Pure Appl. Chem. 61, 517 (1989). 7.
- 8. Molinski, T.F.; Faulkner, D.J.; Tetrahedron Lett. 29, 2137 (1988).
- 9a. Cimino, C.; De Rosa, S.; De Stefano, S.; Spinella, A.; Sodano, G.; Tetrahedron Lett. 25, 2925 (1984).
- b. Loya, S.; Rudi, A.; Tal, R.; Kashman, Y.; Loya, Y.; Hizi, A.; Archiv. of Biochem. and Biophysics, 309, 315 (1994).
- 10. Zeng, C.; Ishibashi, M.; Matsumoto, K.; Nakaika, S.; Kobayashi, J.; Tetrahedron Lett. 37, 8337 $(1993).$
- 11. He Hai-yin, Faulkner, D.J.; J. Org. Chem. 56, 5369 (1991).
- 12. Hvidt, T.; Szarek, W.A.; MaClean, D.B.; Can. J. Chem. 66, 779 (1988).
- 13. Friedländer, P.; Ber. 15, 2572 (1982); Cheng Chia-Chung; Yan Shou-Jen; Org. Reactions, John Wiley & Sons, N.Y. 1982, Vol. 28, chap. 2, p. 37.
- 14. Kempter, G. and Stob, W.; Z. Chem. 2, 61 (1963).
- 15. Tindale, C.R.; Aust. J. Chem. 37, 611 (1984).
- 16. R. Willstätter and A. Phannenstiel; Ber., 37, 4744 (1904).
- 17. a. Inman, W.D.; O'Neill-Johnson, M. and Crews, P.; J. Am. Chem. Soc. 112, 1 (1990). b. West, R.R.; Mayne, C.L. and Ireland, C.M.; Tetrahedron Lett. 31, 3271 (1990).
- 18. Fitzgerald, J.M.; Aust. J. Chem. 16, 246 (1963).
- 19. Omio, S.; Kariyone, K.; Tanaka, K.; Noguchi, M. and Ogiko, T.; Chem. Pharm. Bull. 17, 596 $(1969).$

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